

## CHARACTERIZATION OF MIXTURES OF NATURAL POLYMERIC MATERIAL FOR ENCAPSULATION, BY SPRAY DRYING

### CARACTERIZACIÓN DE MEZCLAS DE MATERIALES POLIMÉRICOS NATURALES PARA ENCAPSULACIÓN, MEDIANTE SECADO POR ASPERSIÓN

**MSc. Luz Alba Caballero Pérez\***, **PhD. Aldo Hernández Monzón\*\***,  
**PhD. Rene Tejedor Arias\*\***, **MSc. Everaldo Joaquin Montes Montes\*\*\***

\* **Universidad de Pamplona**, Facultad de Ingenierías y Arquitecturas. Estudiante de Doctorado en Ciencia y Tecnología de los Alimentos. Universidad de la Habana, Cuba.  
 Directora Grupo de Investigaciones GIBA.

Km 1 Vía Bucaramanga, Campus Universitario, Pamplona, Colombia.  
 Tel.: 057 5685303

E-mail: [luzcaballero@unipamplona.edu.co](mailto:luzcaballero@unipamplona.edu.co)

\*\* **Universidad de la Habana**, Docentes Titulares. Instituto de Farmacia y Alimentos, IFAL. Programa Doctorado en Ciencias de los Alimentos, La Habana, Cuba.  
 E-mail: [renetejedor09@yahoo.es](mailto:renetejedor09@yahoo.es)

\*\*\* **Universidad de Córdoba**, Programa Ingeniería de Alimentos, Montería, Colombia.  
 E-mail: [everaldomontes@yahoo.com](mailto:everaldomontes@yahoo.com)

**How to cite:** Caballero Pérez, L. A., Hernández Monzón, A., Tejedor Arias, R., & Montes Montes, E. J. (2023). CARACTERIZACIÓN DE MEZCLAS DE MATERIALES POLIMÉRICOS NATURALES PARA ENCAPSULACIÓN, MEDIANTE SECADO POR ASPERSIÓN. REVISTA COLOMBIANA DE TECNOLOGÍAS DE AVANZADA (RCTA), 1(41), 1–11.  
<https://doi.org/10.24054/rcta.v1i41.2412>

This work is under an international license  
 Creative Commons Atribución-NoComercial 4.0.



**Abstract:** Background: The characterization of the encapsulating materials to be used in the encapsulation is very important to obtain adequate results and maintain the characteristics of the materials to be encapsulated when subjected to spray drying. Objectives: The objective of this research was to characterize a mixture of natural polymers as an encapsulating material to be used in spray-drying equipment, © Vibrasec brand. Methods: The experimental part was carried out in the facilities of the Engineering Laboratory, University of Córdoba, Montería, Colombia. The physicochemical characterization of the encapsulating material (native cassava starch, oatmeal and sodium alginate) was carried out: granulometric analysis, glass temperature, suspension analysis: pH, acidity, moisture content, apparent density and apparent viscosity, was evaluated. the gelatinization temperature at different concentrations (3.5 and 7%) and temperatures (27, 60 and 90 °C) in order to determine the appropriate temperatures and solid concentrations of the encapsulating material in mixture to prepare the suspensions that meet the drying equipment restrictions (700 mPas) © Vibrasec. Results: The results indicated that sodium alginate should be worked at a maximum of 2% solids concentration and a temperature of 85 °C, while oatmeal (7%) and native cassava starch (10%) should be worked at a temperature of 60°C. In the preparation of the solutions of the mixtures of the encapsulating material, it was established that the maximum concentration of the mixtures should be 12% to obtain a good homogenization of the mixtures and a viscosity (700 mPas) within the restriction of use of the equipment. drying © Vibrasec. Conclusions: It was possible to characterize the encapsulating material made up of oatmeal, sodium

alginate and native cassava starch, establishing the appropriate conditions to be subjected to spray drying in the Marca © Vibrasec equipment.

**Keywords:** sodium alginate, native cassava starch, oatmeal, encapsulating material, spray drying

**Resumen:** Antecedentes: Las actuales tendencias están orientadas al uso de polímeros naturales como material encapsulante en la búsqueda de nuevas alternativas que mantengan la viabilidad de microorganismos a encapsular mediante el secado por aspersión. La caracterización de los materiales encapsulantes a emplear es muy importante para obtener resultados adecuados. Objetivos: El objetivo de esta investigación fue caracterizar una mezcla de polímeros naturales como material encapsulante de una mezcla de cultivos probióticos mediante uso de un equipo de secado por aspersión, marca© Vibrasec. Métodos: La parte experimental se realizó en las Instalaciones del Laboratorio de Ingenierías, Universidad de Córdoba, Montería Colombia. Se realizó la caracterización fisicoquímica del material encapsulante (almidón de yuca nativo, harina de avena y alginato de sodio): análisis granulométrico, temperatura vítre'a, análisis a las suspensiones: pH, acidez, contenido de humedad, densidad aparente y viscosidad aparente, se evaluó la temperatura de gelatinización a diferentes concentraciones (3,5 y 7 %) y temperaturas (27, 60 y 90 °C) con el fin de determinar las temperaturas y concentraciones de sólidos adecuados del material encapsulante en mezcla para preparar las suspensiones que cumplan las restricciones del equipo de secado (700 mPas) © Vibrasec. Resultados: Los resultados indicaron que el alginato de sodio debe trabajarse máximo al 2 % de concentración de sólidos y una temperatura de 85 °C, mientras que la harina de avena (7 %) y el almidón de yuca nativo (10 %) deben trabajarse a una temperatura de 60 °C. En la preparación de las disoluciones de las mezclas del material encapsulante se estableció que la máxima concentración de las mezclas debía ser del 12 % para obtener una buena homogenización de las mezclas y una viscosidad (700 mPas) dentro de la restricción de uso del equipo de secado © Vibrasec. Conclusiones: Se logró caracterizar el material encapsulante conformado por harina de avena, alginato de sodio y almidón de yuca nativo estableciendo las condiciones adecuadas para ser sometidos al secado por aspersión en el equipo Marca © Vibrasec.

**Palabras claves:** alginato de sodio, almidon de yuca nativo, harina de avena, material encapsulante, secado por aspersión.

## 1. INTRODUCTION

Encapsulation is a technique where liquid droplets, solid particles, or gases are covered with a porous polymer film. This membrane, barrier, or film is usually made of components that form a polymer network (Fuchs et al., 2006). The structure created by the encapsulating agent around the encapsulated substance (core) is called a wall; this wall protects the core from deterioration and helps release it under desired conditions (Fuchs et al., 2006, Rodríguez, et al., 2016; Gandomi, et al., 2016; Nunes, et al., 2017).

Spray drying is the most common method of encapsulating food ingredients, as it is more cost-effective compared to other techniques (Arslan, et al., 2015; Eckert, C. et al., 2017), and due to equipment availability and the food stability of the final product (Gul, 2017; Ceja-Medina et al., 2020); This technique can be applied to water-soluble

materials, fish oils (DHA y EPA), natural pigments, probiotic cell concentrates, and powdered milk (Ceja-Medina, et al., 2021; Sun, et al., 2023). In spray drying, the selection of encapsulating agents or coating materials is critical, as they can affect the properties of the emulsion before drying, particle size, and flow properties, as well as the retention of volatile compounds during the process and the shelf life of the powder after drying.

An important factor is the encapsulating agents used, among which we find types, such as lipids: milk fat, lecithins, waxes, stearic acid, monoglycerides, diglycerides, and hydrogenated oils; these are excellent film-formers capable of coating individual particles, providing uniform encapsulation. Carbohydrates are widely used, primarily in spray drying techniques, as encapsulation support. This group includes starches, maltodextrins, and gums. Currently, research is

being conducted on the use of polymeric microencapsulation matrices derived from polysaccharides and their possible combinations to ensure the proper preservation of the encapsulating layer and the active material to be encapsulated (Nie, 2013; Esquivel-González, et al., 2015; Fritzen-Freire, et al., 2021; Madsen, et al., 2022; Mikkel et al., 2022).

The protective material must possess certain properties that depend on the chemical characteristics of the encapsulated material, its application, storage conditions, and the process, as outlined by (Nag, 2011; Moumita, et al., 2017; Homayouni-Rad et al., 2021) he first step in encapsulation is selecting an appropriate encapsulation matrix (Rios-Aguirre, y Gil-Garzon, 2021; Tamtürk, et al., 2023). Research on this topic focuses on the search and application of new coating materials and novel encapsulation techniques (Ávila-Reyes, 2014; Ceja-Medina, et al., 2021; Barros, et al., 2022).

In this research, three materials were evaluated, chosen for their encapsulating and prebiotic properties, and subjected to spray drying using a Vibrasec© brand machine. Native cassava starch (*Manihot sculenta Crantz*) and sodium alginate have been studied for their ability to be used as wall materials for encapsulation, with a wide particle size distribution (Aristizábal, y Sánchez, 2007; Lupo-Pasin et al., 2012; Etchepare, et al., 2016). In the encapsulation of probiotics, these materials are used in moderate concentrations; however, they exhibit low resistance to gastric acids. This characteristic can be counteracted by modifying the gel structure, using mixed polymer systems for matrix formation, or coating the macroparticles (Lupo-Pasin et al., 2012; El-Sayed, et al., 2017; Tao, T., et al., 2019; Triviño, 2019).

Oat flour contains starches,  $\beta$  -  $\alpha$  Glucans and small amounts of proteins, making it a prebiotic material with the potential to form strong, resistant, and flexible films to improve the properties of sodium alginate and native cassava starch (Ta, et al., 2021). The beta-glucan in oats may lead to an FDA-approved health claim (Yonekura, et al., 2014; Yuan, et al., 2023 17). Several studies have published some physicochemical and rheological characteristics of the main components of oats, with notable work conducted by (Berski, et al., 2011; Zamora-Vega, et al., 2012; Zamudio-Flores, et al., 2015; Flores-Peña, et al., 2013).

Research on this topic focuses on the search for and application of new coating materials to be subjected to spray drying. Therefore, this study aimed to evaluate the physicochemical characteristics (solid content and gelatinization temperature) of a mixture of natural polymeric materials with prebiotic properties to encapsulate a mixture of microorganisms using the spray drying technique, in order to determine the appropriate concentration while meeting the viscosity restrictions of the Vibrasec© equipment.

## 2. MATERIALS Y METHODS

The experimental work was conducted at the Nanotechnology Laboratory, Food Technology Center at the University of Pamplona, the Applied Engineering Laboratory, and the Bioprocesses and Fermentations Laboratory at the Berastegui campus of the University of Córdoba, Colombia.

The encapsulating agents used were: Native cassava starch (INNOVAYUCA), sodium alginate (SQ 942), and ground oat flour.

### 2.1. Characterization of encapsulating material and solutions.

The physicochemical evaluation of the encapsulating material and the solutions was carried out as described below:

Table 1: Analysis conducted on encapsulating materials and solutions

Evaluated Parameters	Method Used
Particle size analysis	AOAC. 965.22, (1995)
Moisture content (%)	Gravimetric method by volatilization
Bulk density (g/mL)	Goula, A. M., and Adamopoulos, K. G. (2012)
Glass transition temperature	Paredes-López, O., et al., (1994).
Apparent viscosity of solutions	Brookfield viscometer Nielsen (2010)
Apparent density of solutions	A.O.A.C. 962.37, (1995)
pH	Potentiometric method AOAC 981.12 (2012)
Acidity as lactic acid (% m/m) min	Titration with NaOH AOAC. (942.15, 2005)

Once the solid concentration and glass transition temperature (Tg) of the three encapsulating materials were established, tests were conducted to determine the maximum solid concentration to be used for each material individually and with the mixture of the materials in suspension: heating tests

at different solid concentrations against the apparent viscosity to meet the viscosity requirements for the use of the Vibrasec® equipment, technique employed by (Nielsen, 2010; Aristizábal, y Sánchez, 2007).

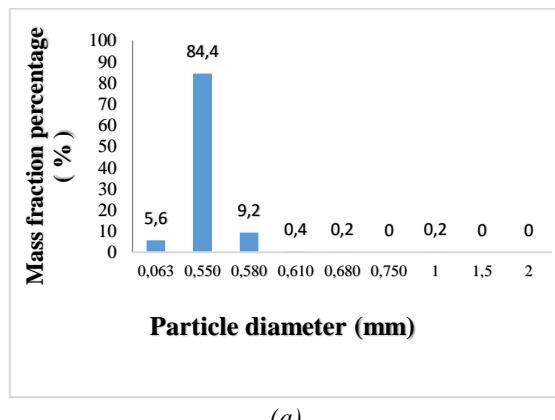
## 2.2 Statistical evaluation of the data

All measurements for the statistical analyses were performed in triplicate. To statistically evaluate each of the experiments conducted, significant differences between treatments were determined by performing an ANOVA analysis with a 95% confidence interval and a Tukey multiple comparisons analysis of the average values.

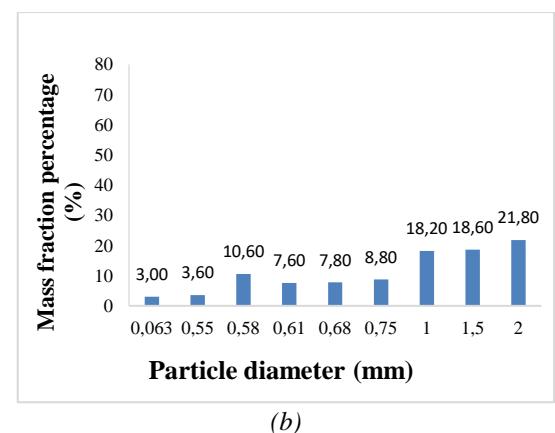
## 3. RESULTS

### 3.1. Particle size analysis of oat flour and native cassava starch

The particle size analysis conducted on the encapsulating materials: oat flour and native cassava starch, allowed for the determination of the particle size distribution, (Flores-Peña, et al., 2013; Aristizábal, y Sánchez, 2007).



(a)



(b)

*Fig. 1. Particle diameter distribution of (a) oat flour and (b) native cassava starch.*

Figure 1 shows the particle diameter distribution of oat flour, with 84.40% having a particle size of 0.55 mm and an average diameter of 0.53 mm, corresponding to a finely ground product (49, 66). The particle size and surface layer of oat flour, due to its high content of dietary fiber, influence the physicochemical properties of the required encapsulating matrix, promoting dissolution and homogenization with the other materials to be mixed. Similarly, the particle size analysis of native cassava starch (*Manihot sculenta Crantz*) is presented, showing a non-homogeneous particle size distribution, where 58.6% fell within the range of 1 to 2 mm and an average interval of 0.54 mm (41.4%), resulting in an average particle diameter of 1.14 mm for cassava starch, which corresponds to a very fine powder (Flores-Peña, et al., 2013; CODEX STAN 176. 1989; De Araujo, et al., 2016).

The results of the particle size analysis of the encapsulating materials show differences in particle size between oat flour and native cassava starch (*Manihot sculenta Crantz*). From fine to very fine, an aspect that was considered for the mixing process, establishing that the preparation of the encapsulating matrix should be performed in a wet manner (dissolution) to prevent the phenomenon of disaggregation, which would hinder mixing. This is in line with what was stated by (Esquivel-González, et al., 2015) who established that a greater number of pores allow for increased particle hydration, and an available surface facilitates the binding of cations; hence the importance of fine granulation in the encapsulating materials to be used.

### 3.2. Physicochemical indicators of the encapsulating materials

Table 2 presents the results of the analyzed indicators:

*Table 2: Physicochemical indicators of the encapsulating materials*

Encapsulating Material N°	Bulk Density (g/mL)	Moisture Content (%)	pH Value ---
Oat flour	1.082 (0.01)	14.67 (0.01)	7.0 (0.01)
Native cassava starch	1.530 (0.03)	12.53 (0.04)	6.5 (0.03)
Sodium alginate	0.990 (0.01)	12.27 (0.07)	7.0 (0.02)

Reported average values correspond to 3 determinations.

As shown in Table 2, the encapsulating materials exhibited pH, bulk density, and moisture content within the ranges of the standards (García-Ceja, and López-Malo, 2012; Madsen, et al., 2022) making them suitable for forming capsules of different sizes due to the molecular interactions of the various polymeric materials to be combined (Barbosa-Cánovas, 2005; Lupo-Pasin, et al., 2012; Arslan, S., Erbas, M., Tontul, I., Y Topuz, 2015).

### 3.3. Differential scanning calorimetry (DSC) of the encapsulating material

Table 3 presents the glass transition temperature ( $T_g$ ) values of the encapsulating material.

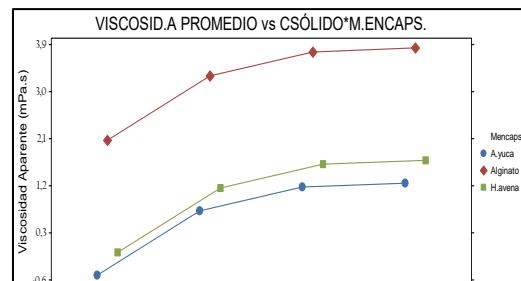
*Table 3: Glass transition temperatures ( $T_g$ ) of the encapsulating materials.*

Encapsulating Material	Glass Transition Temperature, $T_g$ (°C)
Sodium alginate	85
Native cassava starch	63
Oat flour	62

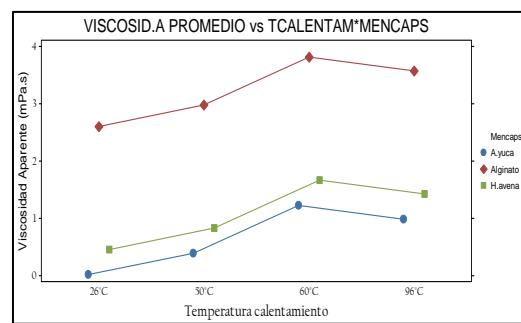
The calorimetry (DSC) results allowed for the evaluation of whether the encapsulating materials to be used exhibited incompatibility within the formulations of both the individual components and the mixture. As shown in Table 3, the glass transition temperature of the three materials is in the range of 62 - 85 °C, which favors obtaining suitable conditions for the solutions when making the different mixtures (Perdomo, et al., 2009; Ceja-Medina, et al., 2020). This result indicates that the feeding temperature of the encapsulating materials should be maintained at temperatures below 63 °C for native cassava starch (*Manihot sculenta Crantz*) and oat flour could be at 60 °C to facilitate its solubilization without gelatinizing (Richardson, et al., 2000), meanwhile, sodium alginate should be processed at a temperature below 85 °C, so they should be prepared separately before mixing.

### 3.4. Apparent viscosity of the encapsulating materials

One of the restrictions of the drying equipment (© Vibrasec) for its use was that the viscosity of the encapsulating materials should not exceed 700 mPa.s. Figure 2 graphically presents the behavior of the apparent viscosity of the three encapsulating materials at the three evaluated concentrations.



(a)



(b)

*Fig. 2. Behavior of the apparent viscosity of the three encapsulating materials (a) versus heating temperature and (b) solid concentration.*

The statistical analysis with  $p$ -values  $< 0.05$  indicates that the apparent viscosity varies according to the encapsulating material, heating temperature, and solid concentration evaluated (see Figure 2).

Table 4 presents the results of the Tukey multiple comparisons analysis of the average values.

*Table 4: Summary of the Tukey multiple comparisons analysis of the average apparent viscosity values*

Statistical technique: ANOVA					
Multiple comparisons test: Tukey					
Significance level: $\alpha = 5\%$					
Statistical software: Statistix versión 10					
Vari ables	Factors	Factor Level	Mean	P- Va lue	Inter preta tion
APPARENT VISCOSITY	Encapsul ating material	Alginate	3,239a		There are differ ences
		Oat flour	1,092b	0.0	
		Cassava starch	0,654c	00	
APPARENT VISCOSITY	Solid concentr ation (%)	7	2,2546a		There are differ ences
		5	2,1796a	0.0	
		3	1,7218b	01	
APPARENT VISCOSITY	Heating temperat ure (°C)	2	0,4923c		There are differ ences
		60	2,2327a		
		50	1,9927ab	0.0	
		96	1,3978b	00	

26	1,0251b	differences
----	---------	-------------

The highest average apparent viscosity was observed with sodium alginate (Nielsen, 2010; Holkem, et al., 2016), followed by native cassava starch and oat flour, with the latter showing the lowest viscosity at the evaluated temperatures and concentrations. Similarly, the average apparent viscosity varies across the three materials depending on the solid concentration. At 7% and 5% concentrations, a statistically similar viscosity is produced between oat flour and native cassava starch (Karimi, et al., 2023) (see Table 4).

On the other hand, the apparent viscosity of the three materials varies according to the heating temperature. It was observed that oat flour and cassava starch exhibited similar viscosities at temperatures between 50 and 60 °C, while sodium alginate showed the highest viscosity values as the temperature and solid concentration increased. Additionally, the highest average viscosity was observed at 60 °C (De Araujo, et al., 2016).

Table 5 shows the results of the viscosity of sodium alginate solutions at temperatures of 60 and 96 °C, considering the water absorption capacity at different solid concentrations of 2% and 3% (w/v), given that the maximum reported value for its use as an encapsulating material for probiotics has been 5% (Rios-Aguirre, et al., 2021).

*Table 5: Apparent viscosity of sodium alginate at different solid concentrations and temperatures.*

Encapsulating agent	Apparent viscosity (mPa.s)			
	Solid concentration/ Temperature (°C)			
	2 / 60	2 / 96	3 / 60	3 / 96
Sodium alginate	404.6 (0.01)	1,040.6 (0.01)	4,296.6 (0.01)	7,490.0 (0.01)

*Reported average values correspond to 3 determinations. p-value <0.05*

The results in Table 5 show that for sodium alginate, the apparent viscosity increased from 404.6 to 7,490 mPa.s in the temperature range of 60 to 96 ± 1 °C, exceeding the maximum allowable value (700 mPa.s) for the use of the drying equipment. Therefore, it was determined that the sodium alginate solution should be prepared at a solid concentration not greater than 2% with constant stirring to facilitate its solubility.

Results corroborating those presented by (Perdomo, et al., 2009; Wan et al., 2010; Mikkel, et al., 2022)

indicate that the viscosity of alginate solutions depends on the concentration. From 2% onwards, the viscosity of the solution increases significantly and exhibits non-Newtonian behavior, meaning its viscosity is not constant.

Table 6 presents the results of the apparent viscosity of native cassava starch and oat flour solutions at concentrations of 3%, 5%, and 7% at a temperature of 60 °C.

*Table 6: Evaluation of the apparent viscosity of native cassava starch and oat flour at different solid concentrations at 60 °C*

Material	Apparent viscosity (mPa.s)		
	3	5	7
Native cassava starch	189.2 (0.07)	356.7 (0.01)	653.7 (0.01)
Oat flour	6.8 (0.00)	10.1 (0.00)	34.5 (0.00)

*Determinations were performed in triplicate, reporting the average value. p-value < 0.05*

As shown in Table 6, the viscosity of native cassava starch increases with increasing concentration and does not exceed the maximum required viscosity value (700 mPa.s); thus, native cassava starch can be used at a maximum concentration of 7% at a temperature of 60 °C. A gradual increase in the apparent viscosity was also observed as the concentration of oat flour in the dispersion increased, with average viscosity values ranging from 6.8 to 34.5 mPa.s, which are below the value set as a restriction. Therefore, oat flour can be used at concentrations above 7% at a temperature of 60 °C.

These results coincide with those reported by Dikeman and Fahey (2006), Lazaridou and Biliaderis (2007), Regand et al. (2011), Rodríguez-Restrepo et al. (2017), and Zamudio-Flores et al. (2021), who indicate that the increases in apparent viscosity values among the dispersions are directly associated with the higher fiber content in the greater concentrations due to the presence of β-glucans in oat flour.

Table 7 shows the apparent viscosity of the suspensions of encapsulating material mixtures at the defined working temperature (60 °C) at different solid concentrations.

*Table 7: Apparent Viscosity of Suspensions of Encapsulating Material Mixtures*

Concentration of solids	Sodium Alginat e	Native Cassava Starch	Oat flou r	Apparent Viscosity
%	%	%	%	mPa.s
16	1	7	8	796 (0.3)
14	1	7	6	693 (0.4)
12	1	5	6	495 (0.6)
10	1	5	4	253 (0.4)

Determinations were performed in triplicate, and the average value is reported.  $p$ -value < 0.05.

As observed in Table 7, for mixture solid values of 16%, the viscosity was above 700 mPa.s, not meeting the restrictions required by the drying equipment. Meanwhile, the mixtures with solid concentrations in the range of 10 to 14% had viscosity values lower than the declared working restriction.

Table 8 shows the behavior of the encapsulating material mixtures subjected to spray drying in the Vibrasec equipment.

**Table 8: Behavior of the Mixture of Materials in Suspension at Different Solid Concentrations in the Vibrasec Equipment**

Solid concentration (%)	Inlet Air Temperature (°C)	Atomization Speed (min <sup>-1</sup> )	Behavior in the Chamber	Moisture Content (%)
20	200	35000	Saturation	---
16	200	35.000	Saturation	9.23
14	200	35.000	Saturation	8.68
12	200	35.000	Normal	4.58
10	200	35.000	Normal	4.32

Based on these results, the mixtures of the encapsulating material were subjected to the spray drying process at a feed temperature of 60 °C and inlet air temperatures of 200 °C. As observed in Table 8, solid concentrations of 14% and 16% exhibited saturation in the chamber and a moisture content between 8.68% and 9.23%. These results align with the findings of Rios-Aguirre et al. (2021), Salinas et al. (2021), and Tamturk et al. (2023), who indicate that the inlet air temperature is directly proportional to the drying rate of the microcapsules and the final water content. Higher adsorbed moisture leads to greater cohesive forces between particles, resulting in increased contact points

among them, which can indirectly affect the dispersion rate of the encapsulating materials, as reflected in the saturation of the drying chamber. In contrast, solid concentrations of 10% and 12% showed normal behavior in the chamber, with moisture content between 4.58% and 4.32%.

This result allowed for the selection of the 12% solid concentration as the most suitable mixture of the encapsulating material in suspension to be used in spray drying under the evaluated conditions of the Vibrasec equipment, discarding the 10% concentration due to its considered low solid content (Soto et al., 2023; Tamturk et al., 2023).

#### 4. CONCLUSIONS.

It is concluded that oat flour, native cassava starch, and sodium alginate possess physicochemical properties that favor their use as encapsulating materials, with the selection of a 12% solid concentration as the most suitable mixture of encapsulating material in suspension to be used in spray drying.

#### CONTRIBUTIONS OF THE AUTHORS

All authors contributed equally to the development and writing of this research.

#### DECLARATION OF COMPETING INTERESTS

The authors declare that they have no conflicts of interest.

#### REFERENCES

- A.O.A.C. 962.37. (1995). Official Methods of Analysis. Association of Official Analytical Chemist. EUA.
- A.O.A.C. 965.22. (1995). Official Methods of Analysis. Association of Official Analytical Chemist. EUA. Análisis granulométrico para harinas.
- A.O.A.C. 981.12. (2012). Official Methods of Analysis. Association of Official Analytical Chemist. EUA. pH.
- AOAC. (942.15, 2005). Official Methods of Analysis. Association of Official Analytical Chemist. EUA. Acidez.

- Aristizábal, J., y Sánchez, T. (2007). Guía técnica para producción y análisis de almidón de yuca. Boletín de servicios agrícolas de la FAO 163. Organización de las Naciones Unidas para la Agricultura y la Alimentación, Roma, Pp. 153.
- Arslan, S., Erbas, M., Tontul, I., Y Topuz, A. (2015). Microencapsulation of probiotic *Saccharomyces cerevisiae* var. *boulardii* with different wall materials by spray drying. *LWT-Food Science and Technology*, 63(1), 685-690.
- Ávila-Reyes, S. V, y otros cuatro autores, (2014). Protection of *L. rhamnosus* by spray-drying using two prebiotics colloids to enhance the viability, doi:<https://doi.org/10.1016/j.carbpol.2013.11.033>, *Carbohydrate Polymers*, 102, 423-432.
- Barbosa-Cánovas, G. V. (2005). Compression and Compaction Characteristics of Selected Food Powders. En Advances in Food and Nutrition doi:10.1016/S1043-4526. Research. Academic Press, Vol. 49 , págs. 233-307.
- Barros, Cp, Silva, R., Guimarães, Jt, Balhtazar, Cf, Verruck, S., Pimentel, Tc, ... & Da Cruz, Ag (2022). Prebióticos y simbióticos en alimentos funcionales. <https://doi.org/10.1002/9781119776345.ch2>. Alimentos funcionales, 21-53.
- Berski, W., A. Ptaszek, P. Ptaszek, R. Ziobro, G. Kowalski, M. Grzesik and B. Achremowicz. (2011). Pasting and rheological properties of oat starch and its derivatives. <https://doi.org/10.1016/j.carbpol.2010.08.036>. *Carbohydrate Polymers* 83.: 665-671.
- Ceja-Medina, L. I., L. y otros siete autores, (2021). In vitro synbiotic activity of *Lactobacillus plantarum* encapsulated with mixtures of Aloe vera mucilage, agave fructans and food additives as wall materials, <https://doi.org/10.24275/rmiq/Bio2234>. *Revista Mexicana de Ingeniería Química*, ISSN-E: 2395-8472, ISSN:1665-2738, 20 (2): 711-723.
- Ceja-Medina, L. I., R. I. Ortiz-Basurto, L. Medina-Torres, F. Calderas, M. J. Bernad-Bernad, R. F. González-Laredo, J. A. Ragazzo-Sánchez, M. Calderón-Santoyo, M. González-Ávila, I. AndradeGonzález & O. Manero. (2020). Microencapsulation of *Lactobacillus plantarum* by spray drying with mixtures of Aloe vera mucilage and agave fructans as wall materials. <https://doi.org/10.1111/jfpe.13436>. *Journal of Food Process Engineering*. 43(8): e13436.
- CODEX STAN 176. (1989). Norma del Codex Para la Harina de Yuca Comestible.
- De Araujo, Scratch Etchepare, M., Raddatz, GC, Cichoski, AJ, Flores, É.M M , Barin , JS , Zepka , LQ , ... & de Menezes , C R. (2016). Efecto del almidón resistente (Hi-maize) sobre la supervivencia de *Lactobacillus acidophilus* microencapsulado con alginato de sodio. *Revista de Alimentos Funcionales*, 21, 321–329.
- Dikeman, C. L. and G. C. Fahey. (2006). Viscosity as related to dietary fiber: A review. Critical <https://doi.org/10.1080/10408390500511862>. *Reviews in Food Science Nutrition* 46 (8):649-663.
- Eckert, C. y otros siete autores, (2017). Microencapsulation of *Lactobacillus plantarum* ATCC 8014 through spray drying and using dairy whey as wall materials, <https://doi.org/10.1016/j.lwt.2017.04.045>, *LWT Food Sci. Technol.* 82, 176–183.
- El-Sayed, H. S., Kassem, J. M., El-Shafei, K. A. W. T. H. E. R., Assem, F. M., & Sharaf, O. M. (2017). Comparative evaluation of the microencapsulation methods efficiency to protect probiotic strains in simulated gastric conditions. *International Journal of Biology, Pharmacy and Allied Science*, ISSN: 2277-4998, 6(3), 521-545.
- Esquivel-González, B., Ochoa Martínez, L., y Rutiaga-Quiñones, O. (2015). Microencapsulación mediante secado por aspersión de compuestos bioactivos. <https://www.redalyc.org/pdf/813/81343176006.pdf>. *Revista Iberoamericana de Tecnología Postcosecha*, 16(2), 180-192.
- Etchepare, M., Raddatz, G. C., Cichoski, A. J., Flores, E. M., Barin, J. S., Queiroz, Z. L., y de Menezes, C. R. (2016). Effect of resistant starch (Himaize) on the survival of *Lactobacillus acidophilus* microencapsulated with sodium alginate. doi:DOI: 10.1016/j.jff.2015.12025. *Journal of Functional Foods*, 21, 321-329.
- Flores-Peña, F. F.; Lozano-Quezada, F.Y.; Ramos-Martínez, A.; Salgado-Delgado, R.; Guerrero-Prieto, V. M.; Ramírez-Mancinas, S.; Bello-Pérez, L. A.; y Zamudio-Flores, P. B. (2013). Caracterización fisicoquímica, reológica y funcional de harina de avena (*Avena sativa* L. cv Bachíniva) cultivada en la región de Cuauhtémoc, Chihuahua. DOI: <https://doi.org/10.54167/tch.v8i3.611>. Revista Tecnociencia. Chihuahua. Vol. VIII, Núm. 3.

- Fritzen-Freire, C. B. et al., (2021). Microencapsulation of bifidobacteria by spray drying in the presence of prebiotics. Available from: Accessed: Nov. 18, doi: 10.1016/j.foodres.2011.09.020, Food Research International, v. 45, p. 306-312.
- Fuchs, M., Turchiuli, C., Bohin, M., Cuvelier, M., Ordonnaud, C., Peyrat-Maillard, M., y Dumoulin, E. (2006). Encapsulation of oil in poder using spray drying and fluidized bed agglomeration. <https://doi.org/10.1016/j.jfoodeng.2005.03.047>. Journal of Food Engineering, 75(1), 27 -35.
- Gandomi, H., y otros cuatro autores, (2016). Effect of chitosan-alginate encapsulation with inulin on survival of *Lactobacillus rhamnosus* GG during apple juice storage and under simulated gastrointestinal conditions, <https://doi.org/10.1016/j.lwt.2016.01.064>.LWT-Food Science and Technology, 69, 365-371.
- García, A., & López, A. (2012). Biopolímeros utilizados en la encapsulación. 6 (1). 84-97. Departamento de Ingeniería Química, Alimentos y Ambiental, Puebla, Recuperado de: <https://www.udlap.mx/wp/tsia/files/No6, 1.>
- Goula, A. M., y Adamopoulos, K. G. (2012). A method for pomegranate seed application in food industries: Seed oil encapsulation,. *Food and Bioproducts Processing*, 90 (4), 639-652. doi:<https://doi.org/10.1016/j.fbp.2012.06.001>.
- Holkem, At, Raddatz, Gc, Nunes, Gl, Cichoski, Aj, Jacob-Lopes, E., Grosso, Crf Y De Menezes, Cr (2016). Desarrollo y caracterización de microcápsulas de alginato que contienen *Bifidobacterium BB-12* producidas por emulsificación/gelificación interna seguida de liofilización. <https://doi.org/10.1016/j.lwt.2016.04.012>. LWT-Ciencia y tecnología de los alimentos,71, 302-308.
- Homayouni-Rad, y otros cuatro autores, (2021). Effect of *Alyssum homolocarpum* mucilage and inulin microencapsulation on the survivability of *Lactobacillus casei* in simulated gastrointestinal and high -temperature conditions, <https://doi.org/10.1016/j.bcab.2021.102075.>, *Biocatalysis and Agricultural Biotechnology* 35: 102075.
- Karimi, R., y otros cuatro autores, (2023). Interaction between β-glucans and gut microbiota: a comprehensive review, <https://doi.org/10.1080/10408398.2023.2192281>, Critical Reviews in Food Science and Nutrition, 1-32.
- Lazaridou, A., & Biliaderis, C. G. (2007). Molecular aspects of cereal β-glucan functionality: Physical properties, technological applications and physiological effects. <https://doi.org/10.1016/j.jcs.2007.05.003>.Journal of cereal science, 46(2), 101-118.
- Lupo-Pasin B, González A. C., Maestro G. A. (2012). Microencapsulación con alginato en alimentos. Técnicas y aplicaciones. *Revista Venezolana de Ciencia y Tecnología de Alimentos.*, 3 (1):(2218-4384), 130-151.
- Madsen, M., y otros cuatro autores, Simulated gastrointestinal digestion of protein alginate complexes: effects of whey protein cross-linking and the composition and degradation of alginate. <https://doi.org/10.1039/D2FO01256A>. *Food & Function*, 13(16), 8375-8387, (2022).
- Mikkel Madsen, Mette E. Rønne, Ruifen Li, Ines Greco, Richard Ipsen y Birte Svensson. (2022). Digestión gastrointestinal simulada de complejos de proteína de alginato: efectos del entrecruzamiento de la proteína de suero y la composición y degradación del alginato<https://doi.org/10.1039/D2FO01256A>. This journal is © The Royal Society of Chemistry. *Food Funct.* volumen13, Pp 8375-8387. N° 16.
- Moumita, S. et al., (2017). Evaluation of the viability of free and encapsulated lactic acid bacteria using in-vitro gastro intestinal model and survivability studies of symbiotic microcapsules in dry food matrix during storage. Doi: 10.1016/j.lwt.2016.11.079. *LWT-Food Science and Technology*, v. 77, p. 40-477.
- Nag, A. (2011). Development of microencapsulation technique for probiotic bacteria *Lactobacillus casei* 431 using a protein polysaccharide complex. New Zealand: Massey University. <http://hdl.handle.net/10179/2355>.
- Nie, S. P., Wang, C., Cui, S. W., Wang, Q., Xie, M. Y., & Phillips, G. O. (2013). A further amendment to the classical core structure of gum Arabic (Acacia senegal). <https://doi.org/10.1016/j.foodhyd.2012.09.014>. *Food Hydrocolloids*, 31(1), 42e48.
- Nielsen, S. (2010). Viscosity Measurement Using a Brookfield Viscometer. Chapter 20. DOI: 10.1007/978-1-4419-1463-7\_20. Food analysis laboratory manual (2), 167 – 168.

- Nunes, G. L. et al., (2017). Inulin, hi-maize, and trehalose as thermal protectants for increasing viability of *Lactobacillus acidophilus* encapsulated by spray drying. Doi: 10.1016/j.lwt.2017.10.032. *LWT- Food Science and Technology*, v. 89, p. 128-133.
- Paredes-López, O., Bello-Pérez, L. A., y López, M. G. (1994). Amylopectin: Structural, gelatinisation and retrogradation studies. Obtenido de [https://doi.org/10.1016/0308-8146\(94\)90215-1](https://doi.org/10.1016/0308-8146(94)90215-1). *Food Chemistry*, 50(4), 411-417.
- Perdomo, J., Cova, A., Sandoval, A., García, L., Laredo, E., y Müller, A. (2009). Glass transition temperature and water sorption isotherms of cassava starch. <https://doi.org/10.1016/j.carbpol.2008.10.023>. *Carbohydrate Polymers*, 76, 305–310.
- Regand, A., Z. Chowdhury, S. M. Tosh, T. M. S. Wolever and P. Wood. (2011). The molecular weight, solubility and viscosity of oat beta-glucan affect human glycemic response by modifying starch digestibility. <https://doi.org/10.1016/j.foodchem.2011.04.053>. *Food Chemistry* 129 : 297-304.
- Rios-Aguirre, Sara y Gil-Garzon, Maritza Andrea. (2021). Microencapsulación por secado por aspersión de compuestos bioactivos en diversas matrices: una revisión. <https://doi.org/10.22430/22565337.1836>. *TecnoL.[online]*. vol.24, n.51, pp.206-229. ISSN 0123-7799.
- Rodríguez, Y.A., Rojas, A.F., Rodríguez-Barona, S. (2016). Encapsulación de probióticos para aplicaciones alimenticias. Doi: 10.17151/biosa.2016.15.2.10. *Revista Biosalud*; 15(2): 106-115.
- Rodríguez-Restrepo, Y. A., Giraldo, G. I., & Rodríguez-Barona, S. (2017). Solubility as a fundamental variable in the characterization of wall material by spray drying of food components: application to microencapsulation of *Bifidobacterium animalis* subsp. *lactis*. <https://doi.org/10.1111/jfpe.12557>. *Journal of Food Process Engineering*, 40(6), e12557.
- Salinas, R. R., Loaiciga, V. Z., & Jaramillo, S. H. (2021). Probióticos: desafíos, revisión y alcance. <https://doi.org/10.31434/rms.v6i6.686>. Revista Médica Sinergia, 6 (6), e686-e686.
- Soto, J. G. M., y otros cinco autores, (2023). Recent developments on wall materials for the microencapsulation of probiotics: A review, DOI: <https://doi.org/10.54167/tch.v17i1.1140>. *Tecnociencia Chihuahua*, 17(1), e1140-e1140.
- Sun, Weizhe, Quang D. Nguyen, Botond Kálmán Süli, Firas Alarawi, Anett Szécsi, Vijai Kumar Gupta, László Ferenc Friedrich, Attila Gere, And Erika Bujna. (2023). Microencapsulación y Aplicación de Bacteria Probiótica *Lactiplantibacillus plantarum* 299v Cepa. *Microorganisms*, 11 (4), 947. <https://doi.org/10.3390/microorganisms11040947>.
- Ta, L. P., y otros seis autores, (2021). Effects of various polysaccharides (alginate, carrageenan, gums, chitosan) and their combination with prebiotic saccharides (resistant starch, lactosucrose, lactulose) on the encapsulation of probiotic bacteria *Lactobacillus casei* 01 strain. *International Journal of Biological Macromolecules*, 183, 1136-1144.
- Tamtürk, F., Gürbüz, B., Toker, Ö. S., Dalabasmaz, S., Malakjani, N., Durmaz, Y., & Konar, N. (2023). Optimization of *Chlorella vulgaris* spray drying using various innovative wall materials. <https://doi.org/10.1016/j.algal.2023.103115>. *Algal Research*, 72, 103115.
- Tao, T., y otros nueve autores, (2019). Influence of polysaccharide as co-encapsulant on powder characteristics, survival and viability of microencapsulated *Lactobacillus paracasei* Lpc-37 by spray drying, <https://doi.org/10.1016/j.jfoodeng.2019.02.009>, *J. Food Eng*, 252, 10-17.
- Triviño Valencia, J. (2019). Efecto almacenamiento y las condiciones de estrés sobre la viabilidad de *bifidobacterium animalis* microencapsulado e incorporado en harina instantánea fortificada a base de plátano dominico hartón (*Musa Aabsimmonds*). Tesis (Magister en Microbiología Agroindustrial). Universidad Católica de Manizales. Instituto de Investigación en Microbiología y Biotecnología Agroindustrial. <https://repositorio.ucm.edu.co/handle/10839/2532>.
- Wang., Q., Hu, X., Du, Y., y Kennedy, J. F. (2010). Alginate/starch blend fibers and their properties for drug controlled release. <https://doi.org/10.1016/j.carbpol.2010.06.004>. *Carbohydrate Polymers*, 82(3), 842-847.

Yonekura, L., Sun, H., Soukoulis, C., Fisk, I. (2014). Microencapsulation of *Lactobacillus acidophilus* NCIMB 701748 in matrices containing soluble fibre by spray drying: Technological characterization, storage stability and survival after in vitro digestión. <https://doi.org/10.1016/j.jff.2013.10.008>. *Journal of Functional Foods*, 6, 205–214.

Yuan, C., y otros cuatro autores, (2023). Extraction and prebiotic potential of  $\beta$ -glucan from highland barley and its application in probiotic microcapsules, <https://doi.org/10.1016/j.foodhyd.>, *Food Hydrocolloids*, 139, 108520.

Zamora-Vega, R., y otros seis autores, (2012), effect of incorporating prebiotics in coating materials for the microencapsulation of *Sacharomyces boulardii*.<https://doi.org/10.3109/09637486.2012.687364>. *International journal of food sciences and nutrition*, 63(8), 930-935.

Zamudio-Flores, P.B. et al. (2015), Digestibilidad in vitro y propiedades térmicas, morfológicas y funcionales de harinas y almidones de avenas de diferentes variedades. *Rev. Mex. Ing. Quím* [online]. vol.14, n.1 [citado 2023-06-06], pp.81-97. Disponible en: <[http://www.scielo.org.mx/scielo.php?script=sci\\_arttext&pid=S1665-27382015000100008&lng=es&nrm=iso](http://www.scielo.org.mx/scielo.php?script=sci_arttext&pid=S1665-27382015000100008&lng=es&nrm=iso)>.

ISSN 1665-2738.